Protocol for Chloroform Methanol Precipitation
For Removal of salt and detergents

1. Add 9x starting volume of iced cold (-20C) Methanol to sample. \*You may need to start in a 15mL or 50 mL to accommodate volumes
2. Vortex well and put in -20 C for at least 30 min
3. Add 1x starting volume of iced cold (-20C) Chloroform to sample
4. Vortex and put in -20 C for another at least 30 min
5. Spin 15 minutes @ 14,0000 g
6. Remove as much solution as possible without disturbing pellet!
7. Add water or TEAB to pellet and bring into solution as best as possible. 200uL would be good. We are next trying to determine weight of protein so as long all is evenly distributed that is OK and volume doesn’t matter just keep the volume as small as possible.
8. You now need to get an accurate mass for each protein
9. Mark an empty Eppendorf tube with proper sample name. Wear gloves at all times when handling tubes.
10. Weigh this dried, empty, marked tube on a scale that has accuracy at the level of 10ug. If you do not have a good scale we strongly recommend that you submit the samples at this point and we will do the measurements.
11. Transfer 10% of your evenly distributed protein solution into this weighed tube and dry down COMPLETELY and re-measure this tube with dried protein pellets. The difference is your protein mass.
12. Once the weight of the 10% fraction is known submit your samples with the accurate mass of each entire precipitation noted on the form.